



Pathology Laboratory Associates

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<https://www.plapath.com>

Bone Marrow Collection Form

Place patient labels here

Place patient labels here

Place patient labels here

Collection Site:

- Right Iliac Crest
- Left Iliac Crest
- Sternum

Ordering Physician: _____

Technician: _____

Bone Marrow Collected by: _____

Indicate the number of specimens collected or received at each location below:

| Specimen Type | Required Quantity | Quantity collected by Client | Quantity received by CPCL Hemo | Quantity received by Processing |
|-------------------------------------|--------------------|------------------------------|--------------------------------|---------------------------------|
| Whole Blood EDTA (Lavender) | 2 tubes | | | |
| BM Aspirate Na Citrate (blue) | 1 tube | | | |
| BM Aspirate EDTA (lavender) | 1 tube | | | |
| BM Aspirate Na Heparin (dark green) | 1 tube | | | |
| BM Clot in Formalin | 1 container | | | |
| BM Biopsy in Formalin | 1 container | | | |
| BM Biopsy in RPMI or Saline | Optional for micro | | | |
| SPS Tube/Micro Isolator | Optional for micro | | | |

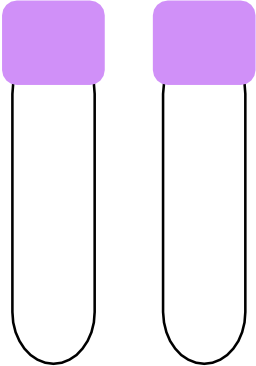
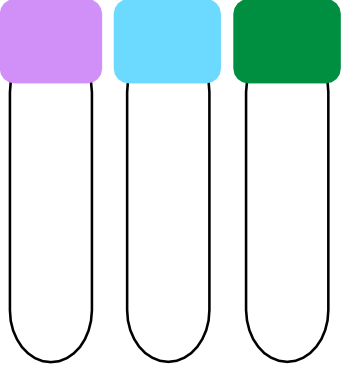
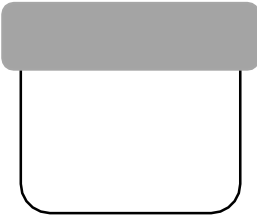
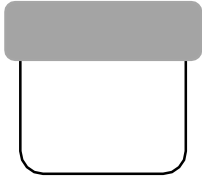
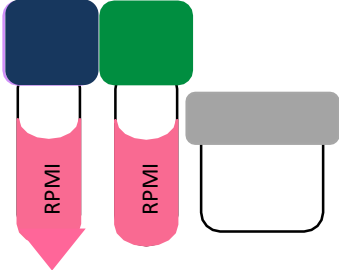
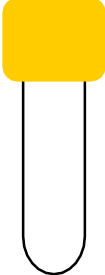
Indicate the number of smears prepared by or received at each location below:
Note: Client preparation of smears is optional, but strongly encouraged.

| Unstained Blood Smear | 2 smears | | | |
|-----------------------------|-------------|--|--|--|
| Stained Blood Smear | Optional | | | |
| Unstained Biopsy Touch Prep | 6-8 preps | | | |
| Unstained BM Smear | 8-10 smears | | | |

Hematology, please make a copy after your portion is completed and return to CPCL Processing to scan into Millennium

Bone Marrow Collection Quick Reference Guide

Every Bone Marrow case should have the following specimens collected:

| Whole Blood | Bone Marrow Aspirate | Bone Marrow Clot | Bone Marrow Core Biopsy | Other |
|--|---|--|--|--|
| <p>2 EDTA Tubes</p>  | <p>1 EDTA Tube + 1 Na Citrate + 1 Na Heparin</p>  <p>(2 mL min per tube)</p> | <p>Formalin Container</p>  | <p>Formalin Container</p>  <p>Dry Tap Collections</p> <p>Prepare 8-10 touch preps, section the biopsy into 3 pieces and place 1 section into each container:</p> <ul style="list-style-type: none"> 1 Conical tube w/RPMI 1 Na Heparin tube w/ RPMI 1 Formalin Container  | <p>Paperwork</p> <ol style="list-style-type: none"> General Requisition Hematopathology Requisition Bone Marrow Collection Form <p>SPS/Isolator Tube (Optional for Microbiology)</p>  |
| <p>Label all specimens with patient name, MRN, DOB, collection date and time AND the specimen type (i.e. blood, aspirate, clot, biopsy)</p> | | | | |



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SPECIMEN COLLECTION:

Each bone marrow should have the following specimens collected:

- Bone marrow aspirate
- Bone marrow clot
- Bone marrow core biopsy
- Peripheral blood

All tubes and containers should be gathered prior to beginning the procedure and labeled with the patient name, MRN, DOB, collection date and time and the specimen types (aspirate, biopsy, clot or blood).

1. First Pull = 3 mL minimum
 - a. Aspirate –
 - i. Place 2 mL of aspirate from the fist pull into a 2.7 mL light blue Sodium Citrate (3%) tube. Mix thoroughly.
Note: ONLY Sodium citrate (3%) is acceptable. Heparin or EDTA anticoagulants should not be used since they may produce artifacts.
 - ii. The remaining 1 mL of bone marrow aspirate is left in the syringe to be used for the clot section.
 - iii. Roll the tube around slowly looking for visible spicule particles. If rare or no visible particles are present, inform collecting personnel so a second aspirate can be attempted. The needle may need to be adjusted or collect a biopsy instead (see dry tap information).
 - iv. Only if making slides at the time of collection, prepare 8-10 pull prep slides from the anticoagulated aspirate. Otherwise, send the aspirate to the lab for preparation of slides. See “slide preparation” below for instructions on how to prepare these slides.
 - b. Clot –
 - i. Allow remaining 1 mL of aspirate to clot in syringe. Slowly remove plunger and clot from syringe.
 - ii. Place clot in a sterile screw top cup containing 10% buffered formalin. Use enough formalin to cover the specimen.
 - iii. Dispose of needle and syringe in a sharps container. Do not transport needles or syringes to the lab.
2. Second Pull = 6-10 mL, minimum 4 mL required
 - a. Distribute the second pull aspiration into the following tubes (2 mL/tube minimum):
 - i. (1) Lavender EDTA tube for Flow Cytometry and Molecular testing
 - ii. (1) Dark green sodium heparin, no gel tube for Chromosome studies
 - iii. (1) Yellow SPS tube, **ONLY** needed if microbiology testing is ordered
 - b. Gently invert each tube several times to allow for proper specimen and anticoagulant mixing.
3. Biopsy
 - a. When collected with Aspirate
 - i. If an aspirate has been obtained, then place the core biopsy in sterile cup containing 10% buffered formalin.
 - ii. **Note:** If possible, prepare 6-8 touch preps using the biopsy prior to placing the specimen in formalin, particularly if the specimen will take several hours to get to the main lab for testing. See instructions below for details on how to prepare these preps.
 - b. Dry tap (No aspirate obtained)
 - i. If there is a dry tap and the physician obtains a small core biopsy, they may attempt to obtain additional biopsy specimens for testing. If one large biopsy is retrieved, then the physician may opt to cut the biopsy into smaller pieces for designated testing, (it is ideal to have at least two pieces).
 - ii. 1° BIOPSY Processing – Touch preparations (6-8 slides) should be made using the biopsy. For Touch prep slide creation, Touch, Do Not press the biopsy to clean slides. Once touch preps are made, place the biopsy in a sterile cup and cover with formalin.
 - iii. 2° BIOPSY Processing- This is either from pieces cut from the initial collection or from a second collection. If from a second collection an additional 6-8 touch preps should be made before processing below.

The biopsy will be sectioned into two separate pieces for FLOW (conical w/ RPMI) and CHROMOSOMES (Na-Heparin w/ RPMI). The core pieces should be covered with RPMI as soon as possible. RPMI must be quality checked prior to addition to specimen.